

The Incidence of ANA and ETI-dsDNA Detected by Enzyme Immunoassays and Indirect Immunofluorescence Assay (IFA)

Izeta Aganovic-Musinovic¹, Lamija Prljaca-Zececic², Djemo Subasic²

¹Center for Genetics, Faculty of Medicine, University of Sarajevo, Bosnia and Herzegovina

²Department for Clinical Immunology, Clinical Center of University of Sarajevo, Bosnia and Herzegovina

ORIGINAL PAPER

SUMMARY

While the SLE (Systemic Lupus Erythematosus) specificity of ANA is low, that of anti-dsDNA autoantibodies is high. The DNA used in the assay must be double stranded: autoantibodies to single –stranded (ss) DNA exist in many diseases and specific to none. The prevalence (70%) of anti-dsDNA autoantibodies is much higher in SLE, giving a higher diagnostic sensitivity than the similarly disease-specific anti-Sm autoantibodies (30%). Anti-dsDNA autoantibodies are usually detected by very analytically sensitive techniques, such

as ELISA (Enzyme Linked Immunosorbent Assay). Within SLE, ds-DNA autoantibodies tend to associate with the presence of glomerulonephritis. Their levels are used to monitor disease activity. We suggest the use of ds-DNA to find the difference between SLE patients with benign variants and classical syndrome of severe skin and renal disease.

Keywords: SLE, systemic lupus erythematosus, ANA, anti-nuclear antibodies, ds-DNA, double-stranded DNA antibodies, IFA, immunofluorescence assay, ELISA, enzyme linked immunosorbent assay

Corresponding author: Izeta Aganovic, MD. Center for Genetics, Faculty of Medicine, University of Sarajevo, Cekalusa 80, 71000 Sarajevo, Bosnia and Herzegovina

1. INTRODUCTION

The sera-immunological hallmark of SLE is anti-nuclear antibody (ANA). In the absence of ANA, the diagnosis of SLE is put into question. So, the ANA is very sensitive test for SLE, being present in virtually all patients and frequently at high titers. Its disease specificity is relatively low since it is frequently found in other rheumatic diseases, as well as in autoimmune liver disease, during viral infection, and, occasionally, at low titers, in normal subjects. It has tendency to increase in prevalence with age in healthy adults.

While the SLE (Systemic Lupus Erythematosus) specificity of ANA is low, that of anti-dsDNA autoantibodies is high. The DNA used in the assay must be double stranded: autoantibodies to single –stranded (ss) DNA exist in many diseases and specific to none

(1). The prevalence (70%) of anti-dsDNA autoantibodies is much higher in SLE, giving a higher diagnostic sensitivity than the similarly disease-specific anti-Sm autoantibodies (30%). Anti-dsDNA autoantibodies are usually detected by very analytically sensitive techniques, such as ELISA (Enzyme linked immunosorbent assay) (1). Within SLE, ds-DNA autoantibodies tend to associate with the presence of glomerulonephritis. Their levels are used to monitor disease activity.

2. MATERIAL AND METHODS

During the period of a year (January 2008-January 2009), we have analyzed 2132 serum specimens using IF method and 1188 sera specimens using ELISA method for ANA and ds-DNA respectively. The most commonly used method for ANA testing is Im-

munofluorescence Assay (2, 3). The basic principle of the procedure is using slides with epithelial cells (Hep-2 cells) as substrate that is incubated in few steps with diluted serum. The unbound material is removed by aspirating and washing. The drop of the fluorescence conjugate (anti human IgG fluorescein labeled containing Blue dye and 0.099 sodium azid) is added (4). Depending on the amounts of autoantibodies in specimens, using IF microscope, it is possible to detect different intensity degree of apple-green fluorescence light. Fluorescence grade is determined as: 4+; 3+; 2+ and +. The ELISA kits are solid-phase enzyme immunoassays. Antigen-precoated microplate wells are incubated with calibrators, controls and serum specimens. During the incubation, antibody present in the test sample binds to the coated wells. Horseradish peroxidase-conjugated anti-human IgG is incubated in the wells to recognize the autoantibodies bound to the coated wells. Chromogen is added and autoantibodies are measured using a spectrophotometer plate reader. At end of each incubation, the unbound material is removed by aspirating and washing. When ANA is considered the positive result value is above 23 while the positive result value for ds-DNA is above 60. Results in range of 20 to 40 are considered negative, while results in range value of 40 to 60 are considered limited positive.

3. RESULTS

During the period of a year we have analyzed 2132 serum specimens using IF method. 924 specimens have been analyzed for ANA antibodies, while 1208 specimens have been analyzed for ds-DNA. All results were interpreted using determined fluorescence grade as 4+; 3+; 2+; + and negative.

Among those 924 specimens analyzed for ANA antibodies the following result were obtained:

- 74% (683) specimens were negative
- 13% (118) specimens were +
- 9% (84) specimens were 2+
- 3% (29) specimens were 3+
- 1% (10) specimens were 4+.

Results are presented on Graph 1.

The same IF method has been used for ds-DNA detection. 1208 specimens were analyzed for ds-DNA antibodies and following results were obtained:

- 94% (1131) specimens were negative
- 2% (25) specimens were +
- 3% (42) specimens were 2+
- 1% (10) specimens were 3+.

Results are presented on Graph 2.

During the same period 1188 serum specimens were analyzed using ELISA method. 745 serum specimens were analyzed for ANA antibodies and results were interpreted as negative if <23 and positive if >23. Results obtained were:

58% (435) specimens were negative
42% (310) specimens were positive

Results are presented on Graph 3.

For ds-DNA using ELISA method were analyzed 443 serum specimens. Results in range of 20 to 40 are considered negative, while results in range value of 40 to 60 are considered limited positive. Results above 60 were positive.

Results obtained were:

- 68% (299) specimens were negative
- 9% (41) specimens were limited positive
- 23% (103) specimens were positive

Results are presented on Graph 4.

4. DISCUSSION

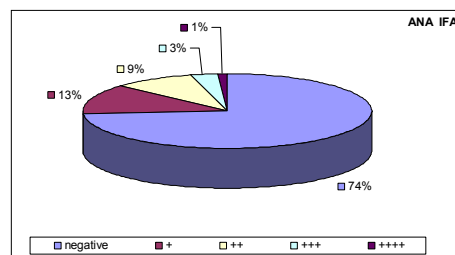
Immunofluorescence assay of ANA showed 74% negative results and 23% positive, either 2+, 3+ or 4+. ELISA tests for ANA antibodies showed 58% negative results and 42% positive results – that in comparing with results achieved using IF – ANA seems to be large number of positive patients, but consider-

ing practicing and quality of ELISA it could be expected. Some studies demonstrated that the commercially available ANA-EIA kits show different levels of sensitivity and specificity (4, 5). On the other hand, at increased temperatures dsDNA denaturizes into ssDNA that results in false negative results using IF method (6, 7). That is the reason why ELISA should be used as confirmation method in those cases. Positive ANA test is not specific for SLE and can be associated with many illnesses as: RA, Sjorgen's syndrome, scleroderma and infectious diseases as mononucleosis, autoimmune thyroid and liver disease. Furthermore, certain medications can cause positive ANA; many healthy people with no expressed illness have a positive ANA (4, 8).

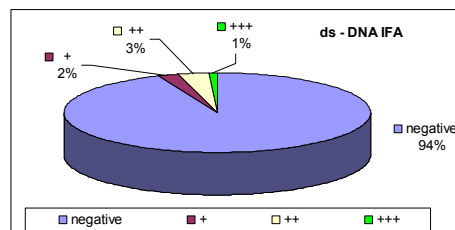
Antibodies to dsDNA (anti-dsDNA, dsDNA-Ab) are frequently found in systemic lupus erythematosus, especially during active disease and differ with respect to immunoglobulin classes and avidity (9). Immunofluorescence assay of dsDNA showed 94% negative specimens and 6% of positive specimens either +, 2+ or 3+. Using ELISA method 68% of specimens were negative, 9% were bordered positive and 23% were positive. Detection of anti-dsDNA may precede the diagnosis of SLE by more than a year. Fluctuations in the level of anti-dsDNA in an individual patient may give important information on the clinical status of the patient (9).

Most of commercial ELISA test systems have great advantages in routine laboratory testing but often detect dsDNA-Ab which are not specific for SLE and therefore give false positive results for non-SLE patients (10, 11).

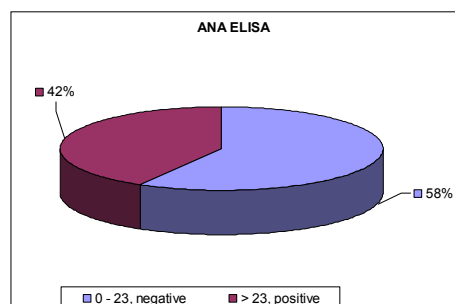
Anti-dsDNA antibody test incorporated in criteria for the classification of SLE needs updating to reflect current insights and technical achievements, including allowance for the presence of non-pathological anti-dsDNA antibodies (12). Anti-DNA of low avidity occurs in rheumatic diseases other than SLE as well, making detection of such antibodies of less diagnostic value (13). It has often been tried to discriminate between clinical subsets of this heterogeneous disease by studying differences within the population of anti-dsDNA antibodies (14). Immunospecificity, complement-fixing ability, avidity, immuno-



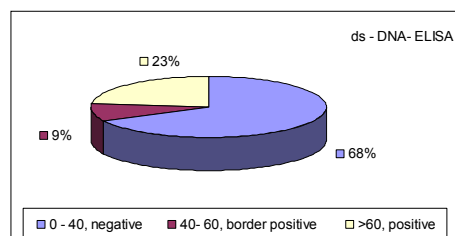
GRAPH 1. Detection of ANA-antibodies using IFA method



GRAPH 2. Detection of Ds-DNA using IFA method



GRAPH 3. Detection of ANA-antibodies using ELISA method



GRAPH 4. Detection of Ds-DNA antibodies using ELISA method

globulin (sub) class composition have all been the subject of different studies, yet conclusions are contradictory and it has not been elucidated (14, 15).

5. CONCLUSION

The most precise method used for SLE detection and keeping up with petition status is ELISA anti-dsDNA, still needs updating. Furthermore, in the presence of antibodies ANA should no longer be considered a valid criterion.

This strategy might ultimately facilitate the difference between SLE patients with benign disease variants and classical syndrome of severe skin and renal disease in pathogenic anti-dsDNA antibodies.

REFERENCES

1. Brinkman K, Termaat R, Van den Brink H. The specificity of the anti-dsDNA ELISA. *J Immunological Methods*, 1991; 139: 91-100.
2. Aarden LA, de Groot ER. and Feltkamp TEW. Immunology of DNA III. Crithidia luciliae, a simple substrate for the determination of anti-dsDNA with the immunofluorescence technique. *Ann NY Acad. Sci*, 1975; 254: 505-15.
3. Stingl G. et al. An immunofluorescence procedure for the demonstration of antibodies to native, double-stranded DNA and circulating DNA-AntiDNA complexes. *Clin Immunol and Immunopathol*, 1976; 6: 131-40.
4. Karamehic J, Subasic D, Gavrankapetanovic F, Zecevic L, Eminovic I, Memic S. et al. The incidence of antinuclear antibodies (ANA) detected by indirect immunofluorescence assay (IFA) method. *Med Arch*, 2007; 61(1): 16-9.
5. Tonuttia E, Bassetti D, Piazza A, Visentini D, Poletto M, Bassetto F. et al. Diagnostic accuracy of ELISA methods as an alternative screening test to indirect immunofluorescence for the detection of antinuclear antibodies. Evaluation of five commercial kits. *Autoimmunity*, 2004; 37(2): 171-6.
6. Grippenberg M, Linder E, Kurki P, Engvall E. A solid phase enzyme-linked immunosorbent assay (ELISA) for the demonstration of antibodies against denatured, single-stranded DNA in patient sera. *Scand J Immunol*, 1978; 7: 151-7.
7. Lange A. Evaluation of the simultaneous estimation of anti-dsDNA and anti-ssDNA antibodies for clinical purposes. *Clin Exp Immunol*, 1978; 31: 472-81.
8. Egner W. The use of laboratory tests in the diagnosis of SLE. *J Clin Pathol*, 2000; 53(6): 424-32.
9. Smeenk RJ, van den Brink HG, Brinkman K, Termaat RM, Barden JH, Swaak AJ. Anti-dsDNA: choice of assay in relation to clinical value. *Rheumatol Int*, 1991; 11(3): 101-7.
10. Villalta D, Romelli PB, Savina C, Bizzaro N, Tozzoli R, Tonutti E. et al. Anti-dsDNA antibody avidity determination by a simple reliable ELISA method for SLE diagnosis and monitoring. *Lupus*, 2003; 12(1): 31-6.
11. Wigand R, Gottschalk R, Falkenbach A, Matthias T, Kaltwasser JP, Hoelzer D. Detection of dsDNA antibodies in diagnosis of systemic lupus erythematosus – comparative studies of diagnostic effectiveness of 3 ELISA methods with different antigens and a Crithidia luciliae immunofluorescence test. *Z Rheumatol*, 1997; 56(2): 53-62.
12. Nossent HC. and Rekvig OP. Is closer linkage between systemic lupus erythematosus and anti-double-stranded antibodies a desirable and attainable? *Arthritis Rees Ther*, 2005; 7(2): 85-7.
13. Smeenk R, Brinkman K, van den Brink H, Termaat RM, Berden J, Nossent H. et al. Antibodies to DNA in patients with systemic lupus erythematosus. Their role in the diagnosis, the follow-up and the pathogenesis of the disease. *Clin Rheumatol*, 1990; 9(1 Suppl. 1): 100-110.
14. Smeenk R, Swaak T. and Smeenk R. Clinical significance of antibodies to double stranded DNA (dsDNA) for systemic lupus erythematosus (SLE). *Clin Rheumatol*, 1987; 6 Suppl. 1: 56-73.
15. Munoz LE, Gaip US, Herrmann M. Predictive value of anti-dsDNA autoantibodies: importance of the assay. *Autoimmun Rev*, 2008; 7(8): 594-7.

Instructions for the authors of the journal Medical Archives

All papers need to be sent electronically by web page: www.avicenapublisher.org. Print version and signed copyright form need to be sent by post to the Editorial board of journal Med Arh. Faculty of Medicine, Cekalusa str. 90, 71000 Sarajevo, BiH. Every sent article gets its number, and author(s) will be notified if their paper is accepted and what is the number of paper. Every correspondence will use that number.

The paper has to be typed on a standard size paper (format A4), leaving left margins to be at least 3 cm. All materials, including tables and references, have to be typed double-spaced, so one page has no more than 2000 alphanumeric characters (30 lines). Sent paper needs to be in the form of triplicate, considering that original one enclosure of the material can be photocopy. Presenting paper depends on its content, but usually it consists of a title page, summary, text references, legends for pictures and pictures.

Title page

Every article has to have a title page with a title of no more than 10 words: name(s), last and first of the author(s), name of the institution the author(s) belongs to, abstract with maximum of 45 letters (including space), footnote with acknowledgments, name of the first author or another person with who correspondence will be maintained.

Summary

The paper needs to contain structured summary (goal, methods, results, discussion, and conclusion) containing up to 300 words, including title, initials of the first name and the last name of the author as well as the name of

the institution. The summary has to contain a list of 3 to 4 keywords.

Central part of the article

Authentic papers contain these parts: introduction, goal, methods, results, discussion and conclusion. Introduction is brief and clear review of problem. Methods are shown so that interested reader is able to repeat described research. Known methods don't need to be identified, it is cited (referenced). If drugs are listed, their genetic name is used (brand name can be written in brackets). Results need to be shown clearly and logically, and their significance proven by statistical analysis. In discussion, results are interpreted and compared to existing, previously published findings in the same field. Conclusions have to give an answer to author's goal.

References

Quoting references must be in a scale in which they are really used. Quoting most recent literature is recommended. Only published articles (or articles accepted for publishing) can be used as references. Not-published observations and personal notifications need to be in text in brackets. Showing references is as how they appear in text. References cited in tables or pictures are also numbered according to quoting order. Citing paper with six or less authors must have cited names of all authors; if seven or more authors' wrote the paper, the name of the first three authors are cited with a note "et al.". If the author is unknown, at the beginning of papers reference, the article is named as "unknown". Titles of the publications are abbreviated in accordance to Index Medicus, but if not listed in the index, whole title of the journal has to be written. Footnote – comments, explanations, etc., cannot be used in the paper.

Statistical analysis

Tests used for statistical analysis need to be shown in text and in tables or pictures containing statistical analysis.

Tables and pictures

Tables have to be numbered and shown by their order, so they can be understood without having to read the paper. Every column needs to have title, every measuring unit (SI) has to be clearly marked, preferably in footnotes below the table, in Arabian numbers or symbols. Pictures also have to be numbered as they appear in text. Drawings need to be enclosed on a white paper or tracing paper, while black and white photo have to be printed on a radiant paper. Legends next to pictures and photos have to be written on a separate A4 format paper. All illustrations (pictures, drawings, diagrams) have to be original and on their backs contain illustration number, first author last name, abbreviated title of the paper and picture top. It is appreciated if author marks the place for table or picture.

Use of abbreviations

Use of abbreviations has to be reduced to minimum. Conventional units can be used without their definitions.

Supplement

If paper contains original contribution to a statistical method or author believes, without quoting original computer program, papers value will be reduced, Editorial staff will consider possibility of publishing mathematical/statistical analysis in-extenso. Papers with the following failure will not be accepted for publishing: grammatically or technically incorrect, materials do not represent original work by author and author(s) have to sign statement that submitted paper has not been published, nor is it currently under consideration for publication elsewhere.